

## Single nucleotide polymorphisms (SNPs) in bovine follicle stimulating hormone receptor (bFSHR) gene in Pasundan cattle

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Bovine follicle stimulating hormone receptor (bFSHR) gene is one of the candidate genes that can be used as a molecular selection for reproductive traits in cattle (Yang et al. 2010). The bFSHR gene plays an important role in follicular development or folliculogenesis (Dierich et al. 1998, Hamny et al. 2017). Activation of FSHR is necessary for the hormonal functioning of FSH and is found at high levels in the ovaries and testes of mammals (Houde et al. 1994). The bFSHR gene of Bos taurus is located in chromosome 11 consisting 10 exons and 11 introns (Simoni et al. 1997, Rahal et al. 1999). Allelic variants in bFSHR gene were reported in several breeds cattle such as Holstein Friesian (Shirasuna et al. 2011, Cory et al. 2012, Arslan et al. 2017), Chinese Holstein (Yang et al. 2010) composite beef cattle (Marson et al. 2008, Hernandez-Cruz et al. 2009) and indigenous Turkish breeds (Arslan et al. 2015).

Previous studies reported that SNP in bFSHR gene located in exon 10 of bFSHR gene (Marson *et al.* 2008, Hernandez-Cruz *et al.* 2009). This SNP can be genotyped using *Alu*I restriction enzyme (ag\*ct) through PCR-RFLP analysis. Several studies reported that this locus (bFSHR/*Alu*I) gene was associated with first ovulation post partum (Shirasuna *et al.* 2011) and number inseminations per pregnancy (Arslan *et al.* 2017). Therefore, the polymorphism of bFSHR/*Alu*I gene was occured in indigenous Turkish cattle (Arslan *et al.* 2015), European-Zebu composite (Marson *et al.* 2008), Holstein cattle (Shirasuna *et al.* 2011), Sudanese cattle (Omer *et al.* 2016) and buffalo (Sosa *et al.* 2015).

There are few literature study that explain the relationship between polymorphism in bFSHR/AluI gene and reproductive traits, included in indigenous Indonesian breeds cattle (Arifin *et al.* 2019). Pasundan cattle is one of Indonesian native cattle from West Java province having limited population. This cattle population was decided to

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Indonesian native cattle based on Ministerial Decree No: 1051/Kpts/SR.120/10/2014. Phenotypic characterization of Pasundan cattle revealed it to be a combination between *Bos javanicus* (Bali cattle) and *Bos indicus* (Ongole and Madura cattle) and developed by crossbreeding from both subspecies in last hundred years (Said *et al.* 2017).

Pasundan cows (77) from the breeding center at Ciamis Regency, West Java (BPPIBT-SP Ciamis) were used for polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP) analysis. 306 bp of PCR products were amplified using primer pairs of Forward: 5'-CTG CCT CCC TCA AGG TGC CCC TC-3' and Reverse: 5'-AGT TCT TGG CTA AAT GTC TTA GGG GG-3' (Houde et al. 1994). The thermal cycling conditions consisted of one cycle of 95°C for 1 min, followed by 35 cycles of 55°C for 15 sec and 72°C for 10 sec with a final extension at 72°C for 5 min. The PCR reaction was performed in 10 µl volume consisting of 5 µl PCR mix (MyTag<sup>TM</sup> HS Red Mix, USA), 0.2 µl of each primer and 4.0 µl of DNA. The PCR products were digested with 0.28 μl of AluI (10 U/μl, 1000 U) restriction enzyme at 37°C for 1 h and separated in 2% agarose gel in 1 × TBE (Tris-Borate-EDTA) buffer and stained with GelRed (Biotium, USA) for visualization under UV light using G-BOX documentation system (Syngene, UK). Sequencing analysis was performed by commercial laboratory service (First BASE Laboratory, Malaysia) to obtain the bFSHR gene in Pasundan cattle.

bFSHR locus in Pasundan cattle is polymorphic with moderate polymorphic informative content (PIC) value (0.30<PIC<0.50) as presented in Table 1. Morever, three genotypes namely CC (243 and 63 bp), GG (193 and 63 bp) and CG (243, 193 and 63 bp) were detected in this syudy as presented in Fig. 1. The frequency of G allele in the bFSHR/AluI of Pasundan cattle was higher than C allele and reveals that GG genotype as the dominant genotype. The moderate PIC value of bFSHR/AluI has been reported in several breeds cattle, i.e. Zebu × British composite breeds (0.37), and 0.34 for indigenous Turkish breeds (Marson et al. 2008, Arslan et al. 2017). Frequency of G allele of bFSHR/AluI locus was higher in Bos taurus, Bos

SNP	Genotypic frequency	Allelic frequency	$H_{e}$	$H_{o}$	n <sub>e</sub>	PIC	$\chi^2$
CC (0.10)	C (0.24)	0.36	0.27	1.57	0.30	52.21	
g.2037C/G	CG (0.27)	G (0.76)					
	GG (0.63)						
	CC (0.02)	C (0.17)	0.28	0.29	1.40	0.24	$0.05^{*}$
g.2071C/T	CT (0.29)	T (0.83)					
	TT (0.68)						
	AA (0.10)	A (0.29)	0.41	0.39	1.71	0.33	$0.14^{*}$
g.2119A/C	AC (0.39)	C (0.71)					
	CC (0.51)						

Table 1. Genetic diversity of three SNPs in the bFSHR gene of Pasundan cattle

SNP, single nucleotide polymorphism;  $H_o$ , observed heterozygosity;  $n_e$ , number of effective allele; PIC, polymorphic informative content; \*under Hardy-Weinberg equilibrium ( $\chi^2$ <5.991).

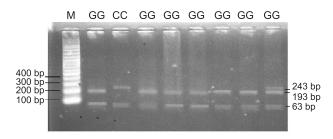


Fig. 1. Detection SNP of g.2037C/G in the bFSHR gene of Pasundan cattle using PCR-RFLP analysis with *Alu*I restriction enzyme showed three genotype namely CC (243 and 63 bp), GG (193 and 63 bp) and CG (243, 193 and 63 bp), M, 100 bp DNA ladder.

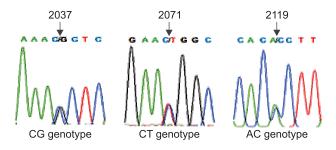


Fig. 2. Partial sequence chromatograms in the exon 10 of bFSHR gene (GenBank: NM\_174061) in Pasundan cattle.

indicus and their crossbreds (Marson et al. 2008, Hernandez-Cruz et al. 2009). bFSHR/AluI locus was monomorphic (CC) in buffaloes (Sosa et al. 2015) and Bos indicus cattle in Sudan, i.e. Butana, Kenana and Erashy (Omer et al. 2016).

Sequencing analysis of partial exon 10 in bFSHR gene from 41 heads of Pasundan cattle revealed two novel single nucleotide polymorphisms (SNPs) at position g. 2071C/T and g.2119A/C (Fig. 2). Milazzotto *et al.* (2000) reported other two novel SNP at position g.1506T/C and g.1539C/T (GenBank: L22319) in bFSHR gene in Nellore cattle (*Bos indicus*).

Present study was conducted to identify reported polymorphism in the exon 10 of bFSHR gene in Pasundan cattle (g.2037C/G) using PCR-RFLP. This SNP had

moderate PIC value (0.30). Sequencing analysis revealed two novel SNPs at g.2071C/T (PIC=0.24) and g.2119A/C (PIC=0.33). Further work for association between bFSHR/ *Alu*I polymorphism and reproductive traits of Pasundan cattle is important.

## REFERENCES

Arifin D N, Said S, Sumantri C, Putra W P B and Hadi D N. 2019. Luteinizing hormone receptor gene polymorphism of Pasundan cattle in Ciamis. *Earth and Environmental Science* 334(2019): 012.

Arslan K, Akyuz B and Agaoglu O K. 2015. Investigation of STAT5A, FSHR and LHR gene polymorphisms in Turkish indigenous cattle breeds (East Anatolian Red, South Anatolian Red, Turkish Grey, Anatolian Black and Zavot). *Russian Journal of Genetics* **51**(11): 1088–95.

Arslan K, Bilal A, Aytac A, Esma G I, Hasan C M and Mehmet U C. 2017. Association of number of artificial inseminations per pregnancy in Holstein dairy cows with polymorphism in Luteinzing Hormone Receptor and Follicle Stimulating Hormone Receptor genes. *Slovenian Veterinary Research* 54(2): 91–98.

Cory A T, Christopher A P, Rejean L and Marie-France P. 2012. Identification of single nucleotide polymorphisms in the bovine follicle-stimulating hormone receptor and effects of genotypes on superovulatory response traits. *Animal Genetics* 44: 197– 201.

Dierich A, Sairam M R, Lucia M, Gian M F, Anne G, Marianne L and Paolo S C. 1998. Impairing Follicle-Stimulating Hormone signaling *in vivo*: Targeted disruption of the FSH receptor leads to aberrant gametogenesis and hormonal imbalance. *Cell Biology* **95**: 13612–17.

Hamny H, Muhammad J, Siti A, Sri W, Widodo and Aulanni'am A. 2017. Polymorphism of follicle stimulating hormone receptor influences the 3D structure and its binding pattern to FSH in *Bos taurus*. *Indian Journal of Animal Research* **51**(4): 630–34.

Hernandez-Cruz B C, Patricia C, Felipe M P, Canseco-Sedano R and Apolo C. 2009. Allelic variants of FSHR gene in cows of different genotypes in Mexico. *Journal of Animal and Veterinary Advances* 8(12): 2489–94.

Houde A, Lambert A, Saumande J, Silversides D W and Lusier J G. 1994. Structure of the bovine follicle-stimulating hormone receptor complementary DNA and expression in bovine tissues. *Molecular Reproduction and Development* **39**(2): 127–

- 35.
- Marson E P, Ferraz J B S, Meirelles F V, Balieiro J C C and Eler J P. 2008. Effect of polymorphisms of LHR and FSHR genes on sexual precocity in a *Bos taurus* × *Bos indicus* beef composite population. *Genetics and Molecular Research* 7(1): 243–51.
- Omer N N, Nahid G, Siham A R and Mohamed-Khair A A. 2016. Genetic characterization of Indigenous Sudanese cattle using FSHR and LHR genes. *American Scientific Research Journal for Engineering, Technology and Sciences* **24**(1): 1–9.
- Rahal P, Latronico A C, Kohek M B F, de Lucia R F S, Milazzotto M P, Wheeler M B, Ferraz J B S, Eler J P and Garcia J F. 1999. Polymorphism in the bovine follicle stimulating hormone receptor gene. *Animal Genetics* 31: 280–91.
- Said S, Widya P B P, Saiful A, Paskah P A and Hani Y. 2017. Phenotypic, morphometric characterization and population structure of Pasundan cattle at West Java, Indonesia. *Biodiversitas* 18(4): 1638–45.

- Shirasuna K, Chiho K, Chiaki M, Yuka A, Yutaka M, Katsuya K, Motozumi M, Takashi S and Akio M. 2011. Relationships between the first ovulation post partum and polymorphism in genes relating to function imunity, metabolism and reproduction in high-producing dairy cows. *Journal of Reproduction and Development* 57(1): 135–42.
- Simoni M, Jorg G and Eberhard N. 1997. The follicle-stimulating hormone receptor: Biochemistry, molecular biology, physiology and pathophysiology. *Endocrine Reviews* 18(6): 739–73.
- Sosa A S A, Karima G M M, Eldebaky H A A, Kandiel M M M, Abou El-Roos M E A and Nawito M F. 2015. *Global Veterinaria* 15(2): 163–68.
- Yang W C, Li S J, Tang K Q, Hua G H, Zhang C Y, Yu J N and Yang L G. 2010. Polymorphisms in the 5'upstream region of the FSH receptor gene and their association with superovulation traits in Chinese Holsteiin cows. *Animal Reproduction Science* 119: 172–77.